

Genetic Polymorphisms Regulating Dopamine Signaling in the Frontal Cortex Interact to Affect Target Detection under High Working Memory Load

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Abstract

■ Frontal-dependent task performance is typically modulated by dopamine (DA) according to an inverted-U pattern, whereby intermediate levels of DA signaling optimizes performance. Numerous studies implicate trait differences in DA signaling based on differences in the catechol-*O*-methyltransferase (COMT) gene in executive function task performance. However, little work has investigated genetic variations in DA signaling downstream from COMT. One candidate is the DA- and cAMP-regulated phosphoprotein of molecular weight 32 kDa (DARPP-32), which mediates signaling through the D1-type DA receptor, the dominant DA receptor in the frontal cortex. Using an *n*-back task, we used signal detection theory to measure performance in a healthy adult population ($n = 97$) genotyped for single nucleotide polymorphisms in the COMT (rs4680) and DARPP-32 (rs907094)

genes. Correct target detection (hits) and false alarms were used to calculate d' measures for each working memory load (0-, 2-, and 3-back). At the highest load (3-back) only, we observed a significant COMT \times DARPP-32 interaction, such that the DARPP-32 T/T genotype enhanced target detection in COMT^{ValVal} individuals, but impaired target detection in COMT^{Met} carriers. These findings suggest that enhanced dopaminergic signaling via the DARPP-32 T allele aids target detection in individuals with presumed low frontal DA (COMT^{ValVal}) but impairs target detection in those with putatively higher frontal DA levels (COMT^{Met} carriers). Moreover, these data support an inverted-U model with intermediate levels of DA signaling optimizing performance on tasks requiring maintenance of mental representations in working memory. ■

INTRODUCTION

A variety of evidence indicates that dopamine (DA) plays a critical neuromodulatory role in the functioning of the PFC (Seamans & Yang, 2004; Goldman-Rakic, 1996). Pharmacological investigation of the primate PFC suggests a critical role for D1-type DA receptor (DRD1) signaling in working memory (WM) tasks (Sawaguchi & Goldman-Rakic, 1991). More recent data from electrophysiological studies of PFC neurons support the notion that DA enhances the signal-to-noise ratio in active PFC networks to enhance signal processing (Kroener, Chandler, Phillips, & Seamans, 2009). More is not always better, however. Converging evidence from animal models supports an “inverted-U” model of PFC DA, whereby an intermediate level of DRD1 stimulation results in optimal WM performance (Williams & Castner, 2006; Goldman-Rakic, Muly, & Williams, 2000), including performance on a delayed response task in rhesus monkeys (Cai & Arnsten, 1997), a delayed version of the radial maze task in rats (Floresco & Phillips, 2001), and a spatial delayed alternation task in mice (Lidow, Koh, & Arnsten, 2003).

Although the tools for investigating the contribution of frontal DA to WM performance in humans are more limited, available evidence supports the view that WM may be similarly regulated in the human PFC. For example, the Val158Met polymorphism (rs4680) in the gene encoding the catechol-*O*-methyltransferase (COMT) enzyme, which regulates PFC DA levels (Kaenmaki et al., 2010; Yavich, Forsberg, Karayiorgou, Gogos, & Mannisto, 2007; Tunbridge, Bannerman, Sharp, & Harrison, 2004; Gogos et al., 1998; Karoum, Chrapusta, & Egan, 1994), has been associated with individual differences in WM performance. Work using the Wisconsin Card Sorting Task (WCST; Malhotra et al., 2002; Egan et al., 2001), which engages a variety of executive processes, including WM, have found that individuals with the COMT Met/Met genotype (and thus higher putative tonic PFC DA) perform better than do Val/Val individuals. Likewise, COMT Met/Met individuals were found to perform better in the 1- and 2-back conditions of a verbal *n*-back WM task relative to Val/Val individuals, but COMT genotype did not predict performance on a continuous performance task that required no updating (Goldberg et al., 2003). The presumed greater persistence of PFC DA in COMT^{Met} allele carriers is proposed to enable more sustained PFC activity and increased

stability of PFC WM representations (Durstewitz, Vittoz, Floresco, & Seamans, 2010; Durstewitz & Seamans, 2008), processes common to the WCST and *n*-back tasks. However, other investigators have failed to replicate an association between COMT genotype and WM performance in humans (Dennis et al., 2010; Bruder et al., 2005). A recent meta-analysis of the role of COMT in cognition suggested that the effect of this polymorphism is minor and that Val homozygotes show higher *n*-back accuracy (Barnett, Scoriels, & Munafo, 2008). These seemingly contradictory findings may be reconciled if one considers that performance on these tasks engages multiple cognitive processes, each of which may require a distinct optimal level or be entirely independent of DA signaling in PFC (Cools & D'Esposito, 2011; Bilder, Volavka, Lachman, & Grace, 2004). Furthermore, growing evidence points to the fact that WM function also depends on DA signaling in the striatum (Rieckmann, Karlsson, Fischer, & Backman, 2011; Stelzel, Basten, Montag, Reuter, & Fiebach, 2010; van Schouwenburg, Aarts, & Cools, 2010; Landau, Lal, O'Neil, Baker, & Jagust, 2009; Cools, Gibbs, Miyakawa, Jagust, & D'Esposito, 2008), where DA is primarily regulated by the DA transporter (DAT), rather than COMT. Thus, considering how COMT may interact with other factors that impact DA signaling may prove useful in understanding the role of frontal DA in WM and other cognitive processes.

As COMT regulates the presynaptic concentration of DA in PFC, we chose to investigate the interaction between COMT genotype and a downstream (postsynaptic) regulator of DA signaling in PFC. As DRD1 receptors are relatively more abundant in the rodent (Gaspar, Bloch, & Le Moine, 1995) and primate PFC than are D2-type DA receptors (Sesack, Snyder, & Lewis, 1995; Goldman-Rakic, Lidow, Smiley, & Williams, 1992; Lidow, Goldman-Rakic, Gallager, & Rakic, 1991) and PFC DRD1 activity is critically involved in WM function (Sawaguchi & Goldman-Rakic, 1991), we chose to investigate a component of the DRD1 signaling cascade with a previously characterized functional polymorphism. Specifically, we investigated a functional single nucleotide polymorphism (SNP; rs907094 A → G/T → C) in the PPP1R1B gene, which encodes the DA- and cAMP-regulated phosphoprotein of molecular weight 32 kDa (DARPP-32; Svenningsson et al., 2004), a downstream mediator of DRD1 stimulation (Nishi, Snyder, & Greengard, 1997; Walaas & Greengard, 1984) thought to play an important role in DA signaling (Greengard, Allen, & Nairn, 1999). DRD1 activation phosphorylates DARPP-32, which ultimately regulates a number of downstream proteins important in neural excitability, including ligand- and voltage-gated ion channels, the sodium/potassium pump, and transcription factors (Greengard et al., 1999). In rodents, DARPP-32 phosphorylation levels in PFC predict performance in PFC-dependent tasks (Kolata et al., 2010; Hotte et al., 2006; Seamans, Floresco, & Phillips, 1998), and DARPP-32 knockout mice demonstrate behavioral deficits suggesting frontal impairment (Heyser, Fienberg, Greengard, & Gold, 2000). Whereas extensive data from

animal models support a role of DARPP-32 in frontal DA-dependent cognitive performance, few studies have investigated DARPP-32 in human cognition. One study found that a DARPP-32 haplotype that includes the rs907094 "A" (or "T") allele is associated with enhanced 3-back performance in an *n*-back task, increased PPP1R1B mRNA expression in PFC (specifically the middle frontal gyrus) of postmortem human brain, and greater structural and functional connectivity between striatum and lateral PFC during *n*-back performance (Meyer-Lindenberg et al., 2007).

Whereas DARPP-32's modulation of DRD1 signaling clearly suggests a role for DARPP-32 in regulating DA signaling in PFC, most work on DARPP-32 to date has focused on its role in modulating striatal function. For example, human studies of the rs907094 SNP have associated it with variations in reinforcement learning (Frank, Doll, Oas-Terpstra, & Moreno, 2009; Frank, Moustafa, Haughey, Curran, & Hutchison, 2007), where it appears to contribute to subtle discrimination abilities (Frank & Fossella, 2011). These findings have been interpreted in terms of DARPP-32 effects in the striatum, where it also modulates DRD2 signaling, albeit with opposing downstream effects: inhibition of cell excitability (Greengard et al., 1999). As a means of specifically isolating cognitive processes in which strong evidence implicates frontal DA and the DRD1 (Vijayraghavan, Wang, Birnbaum, Williams, & Arnsten, 2007), we selected a WM task for which load increases preferentially engage the middle frontal gyrus, an effect that is modulated by putative frontal DA signaling, as indexed by COMT genotype and enzyme activity (Jacobs & D'Esposito, 2011). Distractor-resistant maintenance is thought to rely more specifically on frontal DA, whereas striatal DA appears to primarily subserve successful updating (van Schouwenburg et al., 2010). As load increases boost maintenance demands without markedly changing updating demands, observed effects of DARPP-32 on load-dependent performance would likely reflect mainly maintenance processes, which are largely mediated at the level of PFC.

On the basis of evidence that DARPP-32 potentiates DRD1 signaling and that the rs907094 T/T genotype is associated with enhanced DARPP-32 expression in PFC and enhanced 3-back performance, we hypothesized that this postsynaptic modulator of PFC DA signaling would interact with a presynaptic regulator of PFC DA signaling, the COMT Val158Met SNP, to influence WM performance in humans. Specifically, we expected to see greatest interactive effects of COMT and DARPP-32 genotype on correct target detection under high load in an *n*-back task. We expected this interaction to conform to an inverted-U model of PFC DA signaling, whereby too much or too little DA would impair performance (Cools & D'Esposito, 2011; Goldman-Rakic et al., 2000). To test this hypothesis, we tested healthy adult participants' performance in an *n*-back WM task (Jacobs & D'Esposito, 2011; Gray, Chabris, & Braver, 2003), used signal detection analysis (Macmillan & Creelman, 2005) to investigate target detection in the presence of competing stimuli, and genotyped

participants for the Val158Met COMT and DARPP-32 polymorphisms. We then tested for interacting effects of COMT and DARPP-32 genotype on d' measures of target detection calculated for each WM load.

As DARPP-32 is also known to modulate DA signaling in the striatum (Frank et al., 2007; Meyer-Lindenberg et al., 2007; Ouimet, Miller, Hemmings, Walaas, & Greengard, 1984), we also tested whether replacing DARPP-32 genotype in our statistical models with a genetic marker of striatal DA tone could produce similar results. To do so, we genotyped participants for a variable number tandem repeat (VNTR) polymorphism in the 3' untranslated region of the DAT gene (SLC6A3; Vandenberg et al., 1992). This choice was based on evidence that DAT is the primary mechanism of DA clearance in the striatum (Lewis et al., 2001; Jones, Gainetdinov, Wightman, & Caron, 1998; Sesack, Hawrylak, Matus, Guido, & Levey, 1998; Giros, Jaber, Jones, Wightman, & Caron, 1996) and that variations in the number of repeats in this VNTR affect DAT availability in the striatum, with the most support for increased DAT availability in 9 repeat (9R) carriers (Shumay, Chen, Fowler, & Volkow, 2011; van de Giessen et al., 2009; but cf. VanNess, Owens, & Kilts, 2005; Heinz et al., 2000).

METHODS

Participants

Participants ($n = 97$) were recruited from the University of North Carolina, Chapel Hill (UNC) and surrounding community. All participants were healthy individuals aged 22–40 years old with no history of neurological or psychiatric disorders and no current use of psychoactive medications or other psychoactive substances aside from moderate caffeine, nicotine, or alcohol. Participants were native English speakers, had at least a high-school education, and reported lifetime alcohol consumption of ≥ 1 drink. We excluded people under 22 based on the finding that age modulates COMT genotype effects on another aspect of cognition (Smith & Boettiger, 2012). This age cutoff is also supported by data showing that brain maturation asymptotes at ~ 22 years (Dosenbach et al., 2010). We collected information regarding participants' personal and parental occupation and education via questionnaire to quantify socioeconomic status (SES; Hollingshead, 1975). Participants gave written informed consent, as approved by the UNC Office of Human Research Ethics. Participants received monetary compensation for participating.

Genotyping

COMT and DARPP-32 Single-Nucleotide Polymorphisms

DNA samples were extracted from saliva samples using Oragene DNA Collection Kits (DNAGenotek, Kanata, Ontario, Canada). DNA samples from each participant were genotyped for the Val158Met COMT (rs4680) and DARPP-32 (rs907094) polymorphisms using TaqMan

technology (Applied Biosystems, Foster City, CA) as described previously (Smith & Boettiger, 2012; Boettiger et al., 2007). Genotyping was performed by the Duke Center for Human Genetics. Both genotypes were in Hardy-Weinberg equilibrium (COMT: $\chi^2 = 1.23$, $df = 2$, $p = .54$; DARPP-32: $\chi^2 = 0.785$, $df = 2$, $p = .675$). The COMT genotype distribution was 29:43:25 (Val/Val:Val/Met:Met/Met). The DARPP-32 genotype distribution was 9:35:53 (C/C:C/T:T/T). The C/C and C/T genotypes were combined as C-carriers for all analyses to achieve comparable group sizes. COMT distributions did not differ between DARPP-32 groups, $\chi^2 = 1.38$, $df = 2$, $p = .5$.

DAT 3' UTR VNTR Polymorphism

DNA samples were also genotyped for the DAT 3' UTR VNTR using a modification of a previously described protocol (Anchordoquy, McGeary, Liu, Krauter, & Smolen, 2003). The PCR primer sequences were 5'-TGT GGT GTA GGG AAC GGC CTG AG-3' and 5'-CTT CCT GGA GGT CAC GGC TCA AGG-3'.

PCR was carried out in a 20- μ l final reaction volume with 1.0 U AmpliTaq Gold polymerase (Life Technologies, Foster City, CA), 1 \times PCR buffer II solution (Life Technologies, Foster City, CA), 10% dimethyl sulfoxide (DMSO; Hybra-Max grade; Sigma, St. Louis, MO), 180 μ M dNTPs, 20 ng of DNA, with 7'-deaza-2'-deoxy-GTP (Roche Applied Science, Indianapolis, IN) substituted for one half of the dGTP, and 480 nM for forward and reverse DAT primers (Life Technologies, Foster City, CA). Cycling conditions were as follows: 95°C for 10 min, with a series of touch down reactions for two cycles at 95°C for 30 sec, 65°C for 30 sec, 72°C for 1 min, two cycles each with a 65°C, 63°C, 61°C, 59°C, and 57°C annealing, respectively, followed by 30 cycles of 95°C for 30 sec, 55°C for 30 sec, and 72°C for 1 min. Final extension was at 72°C for 30 min.

After DAT VNTR amplification, all DNA samples were read for base pair (bp) length by the Genome Analysis facility at UNC using an ABI 3730xl DNA Analyzer (ABI, Foster City, CA) to determine the number of 40 bp repeats in the 3' UTR region of the DAT gene. Data were visualized using PeakScanner software (ABI, Foster City, CA), and base pair calls were made by two independent observers. Known DAT VNTR control samples were provided by Andrew Smolen (University of Colorado, Boulder) and ABI (CEPH 1347-02) for quality assurance of PCR reaction and VNTR readout. We ascertained DAT genotypes for 96 participants. The distribution of DAT genotypes was 54:29:11:2 (10/10:9/10:9/9:8/10), which is consistent with published data (Mitchell et al., 2000; Kang, Palmatier, & Kidd, 1999; Vandenberg et al., 1992). Participants with rare alleles were excluded from our analyses ($n = 2$).

n-back Task

Our behavioral paradigm was based on a previously described task (Jacobs & D'Esposito, 2011; Gray et al.,

2003). Briefly, the task included 16 blocks of differing WM load (eight 0-back blocks, four 2-back blocks, and four 3-back blocks) ordered in one of two pseudorandom sequences, which were assigned alternatively to each participant (i.e., odd subject numbers received one block sequence, whereas even subject numbers received the same trials presented in the other block sequence). Participants were instructed on the condition (0-, 2-, or 3-back) at the start of each block. A block consisted of 20 serially presented consonant letter stimuli (duration = 1 sec), with each letter followed by a 1-sec delay. 0-back blocks consisted of 20% targets and 80% nonlures, whereas 2- and 3-back blocks consisted of 20% targets, 15% lures, and 65% nonlures. For 0-back trials, the target letter was defined as the letter “X,” whereas in 2- and 3-back trials, target letters were those that matched the letter seen 2 or 3 previously in the stream, respectively. Participants were asked to press one button for targets and another button for nontargets. Lures were letters seen previously in the stream but not at the target position; these were always ± 1 position from the target position. Accuracy and RT measures were collected for each trial.

Data Analysis

Discriminating Targets from Nontargets

To quantify participants’ ability to correctly detect targets from the stimulus stream, we calculated a target discriminability index. The index, d' , is the most commonly employed index derived from signal detection theory (Green & Swets, 1966). Proportion of hits (correct target identification) and false alarms (responding to nontarget nonlures and/or lures as targets) were calculated for 0-, 2-, and 3-back conditions and used to calculate a d' measure of target detection for each WM load. We calculated d' for each participant at each load as follows:

$$d' = Z \text{ transform } (p(\text{Hits})) - Z \text{ transform } (p(\text{False Alarms}))$$

where $p(X)$ = proportion of X instances (Hits or False Alarms) across all trials of a common load (0-, 2-, or 3-back). Hits reflected correct identification of a target as a target. The maximum Hit probability was set at 31/32 (0.969, Z score = 1.863) for 0-back trials and 15/16 (0.938, Z score = 1.534) for 2- and 3-back trials to allow for Z -transform computation. Incorrect identification of any nontarget (nonlure or lure) as a target was classified as a False Alarm. The minimum False Alarm probability was set at 1/128 (0.0078, Z score = -2.418) for 0-back trials and 1/64 (0.0156, Z score = -2.154) for 2- and 3-back trials. Our Z transform reflects the Z -score transform of the proportion of responses based on a normal distribution of response probabilities mean centered at a probability of 0.5 (Z score = 0).

Discriminating Targets from Lures

For the 2- and 3-back conditions, a separate d' measure was calculated based on target hits versus lure false alarms as follows:

$$d' = Z \text{ transform } (p(\text{Hits})) - Z \text{ transform } (p(\text{Lure False Alarms}))$$

where Lure False Alarms reflect incorrect identification of lures as targets. The maximum Hit probability was set at 15/16 (0.938, Z score = 1.534) for 2- and 3-back trials, as for the d' calculation above. The minimum Lure False Alarm probability was set at 1/12 (0.0833, Z score = -1.383) for 2- and 3-back trials.

d' Slope

The use of different WM load levels allowed us to calculate the d' slope (the difference in d' between the 2- and 3-back conditions divided by the load difference). This measure quantifies the effect of increasing WM load on performance per unit of memoranda.

Statistical Analyses

To test the significance of across-group comparisons, we used 3 (COMT group: Met/Met, Val/Met, Val/Val individuals) \times 2 (DARPP-32 group: C carriers, T/T individuals) ANOVA for continuous measures and χ^2 tests for categorical measures. Repeated-measures ANOVA with Genotype Group (3 \times 2) as between-subject factors and WM Load as a within-subject factor (3 levels) were performed on accuracy and RT data. Analyses including DAT data were identical, with the substitution of DAT genotype (9R carrier, 10/10 individuals), for DARPP-32 genotype. All ANOVA included participant sex and ethnicity as covariates (see Demographic data in Results for justification). When necessary, a Greenhouse–Geisser nonsphericity correction was applied. Post hoc comparisons were performed where indicated using ANOVA and two-tailed t tests. All statistical procedures were carried out in SPSS (SPSS Inc., Chicago, IL). Values reported as mean \pm SEM, unless otherwise stated.

RESULTS

Demographic Data

Demographic factors, including years of education, SES (Hollingshead, 1975), and age, did not significantly vary as a function of either COMT or DARPP-32 genotype in our sample (Table 1). Furthermore, gender ratios were well balanced across groups. Although a greater proportion of DARPP-32 T/T individuals were white, this was not unexpected, given that allelic distribution of this polymorphism is known to vary as a function of ethnicity, with T being the major allele in white and Japanese populations and

Table 1. Demographic Data by Genotype Groups

	<i>COMT</i> V/V <i>D-32 C</i> (<i>n</i> = 15)	<i>COMT</i> V/M <i>D-32 C</i> (<i>n</i> = 20)	<i>COMT</i> M/M <i>D-32 C</i> (<i>n</i> = 9)	<i>COMT</i> V/V <i>D-32 T/T</i> (<i>n</i> = 14)	<i>COMT</i> V/M <i>D-32 T/T</i> (<i>n</i> = 23)	<i>COMT</i> M/M <i>D-32 T/T</i> (<i>n</i> = 16)	<i>F</i> (2, 91) (<i>p</i>)
Age (years)	25.5 ± 4.2	26.7 ± 5.6	25.9 ± 4.2	24.8 ± 4.0	24.8 ± 4.7	25.1 ± 4.6	0.18 (.84)
Edu (years)	16.6 ± 1.5	16.4 ± 1.4	17.0 ± 2.4	16.8 ± 2.3	16.7 ± 1.6	16.1 ± 0.8	0.96 (.39)
Participant SES	44.1 ± 5.3	46.1 ± 6.0	48.3 ± 4.0	46.1 ± 6.2	47.2 ± 6.3	46.1 ± 5.8	0.89 (.42)
Sex (% female)	53.3	50.0	77.8	35.7	60.9	68.8	(.34) ^a
Ethnicity (% White)	40	55.0	66.7	92.9	91.3	87.5	(.001) ^a

Values are reported as mean ± standard deviation. Reported *p* values reflect *COMT* × *DARPP-32* interaction effects on the variable of interest following 3 (*COMT*) × 2 (*DARPP-32*) ANOVAs. Exact *p* values reported unless *p* < .001. D-32 C = carrier of *DARPP-32 C* allele; D-32 T/T = *DARPP-32 T/T* individuals; Edu = education; V = valine; M = methionine.

^a*p* value represents results of χ^2 test.

C being the major allele in an African population (Yoruba, Nigeria; Frazer et al., 2007). To control for this potential confound of ethnic distribution differences across our genotype groups, ethnicity was included as a covariate in our analyses. As indicated in the results, we also repeated our ANOVA and post hoc testing within the white participants (*n* = 71) to verify that any significant *DARPP-32* × *COMT* interaction effects in the full sample were not driven by differences in ethnic distribution between genotype groups. We also included sex as a covariate in our analyses, based on data showing that sex moderates *COMT* activity in postmortem human PFC (Chen et al., 2004).

Overall *n*-Back Performance Declines as a Function of Load

The load manipulation produced significant differences in both Target accuracy, $F(2, 178) = 6.97, p = .001$, and RT to correctly identify Targets, $F(2, 178) = 23.21, p < .001$.

Load effects on accuracy reflected near significant decreases in Target accuracy between 0- and 2-back trials, $F(1, 89) = 3.69, p = .058$, and between 2- and 3-back trials, $F(1, 89) = 3.57, p = .062$. Load effects on RT for correct Target identification reflect significant slowing from 0- and 2-back trials, $F(1, 89) = 20.7, p < .001$, and from 2- to 3-back trials, $F(1, 89) = 5.00, p = .03$. We observed no significant main or interacting effects of *COMT* or *DARPP-32* genotype on Target accuracy or RT (max $F = 2.68$, min $p = .07$, main effect of *COMT* on target accuracy; Table 2).

Target versus Nontarget Discrimination Decreases with Increasing Load

Beyond simple performance estimates, we wished to determine how well participants were able to discriminate targets imbedded within a stream of nontargets. To do so, we used signal detection theory to calculate d' as a

Table 2. *n*-Back Task Performance for Target Stimuli by Genotype Groups

	<i>COMT</i> V/V <i>D-32 C</i> (<i>n</i> = 15)	<i>COMT</i> V/M <i>D-32 C</i> (<i>n</i> = 20)	<i>COMT</i> M/M <i>D-32 C</i> (<i>n</i> = 9)	<i>COMT</i> V/V <i>D-32 T/T</i> (<i>n</i> = 14)	<i>COMT</i> V/M <i>D-32 T/T</i> (<i>n</i> = 23)	<i>COMT</i> M/M <i>D-32 T/T</i> (<i>n</i> = 16)
<i>Target Accuracy (%)</i>						
0-back	92.9 ± 2.0	90.7 ± 1.6	85.2 ± 2.4	92.9 ± 2.0	89.6 ± 1.5	92.9 ± 1.8
2-back	88.8 ± 4.2	86.0 ± 3.5	70.9 ± 5.2	82.4 ± 4.3	81.4 ± 3.3	77.2 ± 3.9
3-back	66.0 ± 4.1	69.0 ± 3.5	62.5 ± 5.1	70.2 ± 4.1	61.7 ± 3.2	66.5 ± 3.8
<i>Target Correct RT (msec)</i>						
0-back	481.1 ± 18.1	497.5 ± 15.2	507.8 ± 22.4	516.1 ± 18.3	494.8 ± 14.2	537.5 ± 16.9
2-back	690.1 ± 32.1	685.0 ± 27.0	724.4 ± 39.6	710.9 ± 32.4	699.1 ± 25.2	707.6 ± 29.9
3-back	729.0 ± 33.6	758.5 ± 28.3	724.2 ± 41.5	771.8 ± 33.9	726.2 ± 26.4	793.3 ± 31.4

Values are reported as mean ± SEM. Conventions as for Table 1.

Table 3. Basic Target Discrimination across WM Loads by Genotype Groups

	COMT V/V D-32 C (n = 15)	COMT V/M D-32 C (n = 20)	COMT M/M D-32 C (n = 9)	COMT V/V D-32 T/T (n = 14)	COMT V/M D-32 T/T (n = 23)	COMT M/M D-32 T/T (n = 16)
0-back	3.87 ± 0.11	3.76 ± 0.09	3.47 ± 0.14	3.93 ± 0.11	3.70 ± 0.09	3.86 ± 0.10
2-back	2.76 ± 0.21	2.85 ± 0.17	2.03 ± 0.25	2.58 ± 0.20	2.70 ± 0.16	2.50 ± 0.19
3-back	1.86 ± 0.15	1.95 ± 0.12	1.70 ± 0.18	2.09 ± 0.15	1.70 ± 0.12	1.85 ± 0.14

Target discrimination quantified as a d' measure of correct target hits versus nontarget false alarms (see Methods). Values are reported as mean ± standard error. Other conventions as for Tables 1 and 2. Significant main effect of WM load: $F(2, 178) = 22.58, p < .001$; no significant main or interacting effects of COMT or DARPP-32 genotype on basic target detection (max $F = 1.69$, min $p = .19$).

discriminability index (see Methods). We performed this analysis first including all trial types (Table 3). A repeated-measures ANOVA (Load × COMT × DARPP-32) taking d' as the dependent measure found a significant main effect of WM Load on d' , $F(2, 178) = 22.58, p < .001$. This reflected significant decreases in d' from both the 0- (3.76 ± 0.04) to 2-back condition (2.57 ± 0.08; $F(1, 89) = 13.15, p < .001$) and the 2- to 3-back condition (1.86 ± 0.06; $F(1, 89) = 8.35, p = .005$). In contrast, we observed no significant main effect of COMT, $F(2, 89) = 2.8, p = .066$, or DARPP-32, $F(1, 89) = 0.59, p = .45$, nor any significant COMT × DARPP-32 interaction, $F(2, 89) = 2.45, p = .093$, on basic target detection.

Discrimination of Targets from Lures

As previous work with this particular n -back paradigm found an inverted-U-shaped relationship between DA signaling and both WM performance and neural activity when considering lure trials (Jacobs & D'Esposito, 2011), we investigated the relationship between COMT and DARPP-32 genotypes on discrimination between target and lures for 2- and 3-back trials (target/lure d' ; see Table S1 for Lure RT and accuracy data by genotype group). A 3 (COMT) × 2 (DARPP-32) × 2 (WM load) mixed repeated-measures ANOVA with Target/Lure d' as the dependent measure found a significant effect of WM load on d' , $F(1, 89) = 8.98, p = .004$, with increasing WM maintenance demands from the 2- to 3-back condition decreasing d' from 1.65 ± 0.09 to 0.93 ± 0.07, respectively. We did not observe significant main effects of either COMT, $F(2, 89) = 1.41, p = .25$, or DARPP-32 genotype, $F(1, 89) = 0.028, p = .87$, on overall target/lure discrimination. Critically, we did find a significant COMT × DARPP-32 × Load interaction effect on target versus lure discrimination, $F(2, 89) = 4.61, p = .012$. Given our somewhat ethnically mixed sample (see Table 1) and reported ethnic differences in $COMT^{Val^{158}Met}$ allele frequencies (e.g., McLeod et al., 1998; McLeod, Fang, Luo, Scott, & Evans, 1994), it is important to note that the COMT genotype × DARPP-32 genotype × WM load interaction remained nearly significant when our analyses were restricted to the white participants, the largest ethnic group included, $F(2, 64) = 3.09, p = .052$, despite the sub-

stantial loss of statistical power. We also found a significant COMT × Load effect, $F(2, 89) = 6.03, p = .003$, but no DARPP-32 × Load effect, $F(1, 89) = 0.36, p = .55$, on target/lure d' .

We next performed post hoc ANOVA to investigate the COMT × DARPP-32 effect on target/lure discrimination separately for the 2- and 3-back conditions. Although we observed a main effect of COMT for target/lure discrimination in 2-back trials, $F(2, 89) = 3.97, p = .022$, no main effect of DARPP-32 or interaction effects were observed (max $F = 1.37$, min $p = .26$). In contrast, for 3-back trials, we found a significant COMT × DARPP-32 interaction, $F(2, 89) = 4.37, p = .016$ (Figure 1), with no main effect of either COMT, $F(2, 89) = 0.45, p = .64$, or DARPP-32, $F(1, 89) = 0.03, p = .86$. This COMT × DARPP-32 interaction effect on 3-back target/lure d' also remained when only white participants were considered,

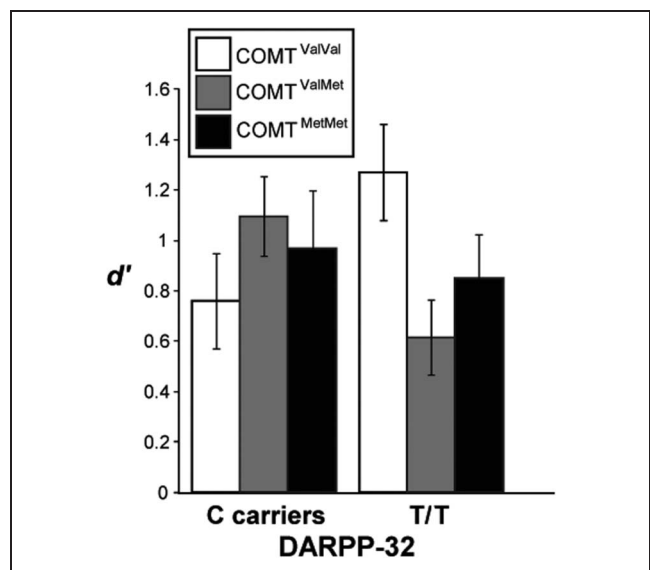


Figure 1. COMT and DARPP-32 genotype interaction effects on WM performance. Graphs depict target discrimination (d' for targets versus lures; mean ± SEM). Under high WM load (3-back trials), COMT and DARPP-32 genotype significantly interact to affect d' , according to an inverted-U relationship, $F(2, 89) = 4.37, p = .016$. A post hoc ANOVA found a significant effect of DARPP-32 genotype on 3-back target/lure detection in $COMT^{ValMet}$ individuals, $F(1, 39) = 6.23, p = .017$.

$F(2, 64) = 4.05, p = .022$. This interaction primarily reflects impaired target/lure discrimination in COMT^{Val/Met} individuals with the DARPP-32 T/T genotype, $F(1, 39) = 6.23, p = .017$ (Figure 1). In contrast, the DARPP-32 T/T genotype was associated with enhanced target/lure discrimination in COMT^{Val/Val} individuals, although the latter effect failed to reach statistical significance, $F(1, 25) = 2.14, p = .156$ (Figure 1). No significant effects of DARPP-32 genotype on 3-back target/lure discrimination were observed among COMT^{Met/Met} individuals, $F(1, 21) = 0.058, p = .81$. When restricting our post hoc analyses to White participants, the DARPP-32 T/T genotype remained associated with impaired target/lure discrimination in COMT^{Val/Met} individuals, $F(1, 29) = 4.65, p = .04$, and we observed a trend toward enhanced target/lure discrimination in COMT^{Val/Val} individuals, $F(1, 16) = 3.64, p = .074$; again, no effect of DARPP-32 was observed among COMT^{Met/Met} individuals, $F(1, 17) = 0.06, p = .81$.

We directly compared the effect of DARPP-32 genotype in the COMT^{Val/Met} group relative to the COMT^{Val/Val} and COMT^{Met/Met} groups. The COMT^{Val/Met} group differed significantly from the COMT^{Val/Val} group on 3-back target/lure discrimination, $F(1, 66) = 8.31, p = .005$, but did not differ from the COMT^{Met/Met} group, $F(1, 62) = 1.2, p = .28$.

To test for interacting effects of DARPP-32 and COMT genotypes on target/lure discrimination with increasing load, controlling for baseline WM performance, we next evaluated the d' slope for target/lure discrimination. In addition to normalizing high WM load (3-back) performance by baseline WM (2-back) performance, the d' slope quantifies the change in target detection per unit of memorandum in WM and can be more readily compared across studies with differing load levels. More negative slopes indicate poorer target discrimination as a function of WM load. A 3×2 ANOVA (COMT \times DARPP-32) found a significant COMT \times DARPP-32 interaction, $F(2, 89) = 4.26, p = .017$ (Figure 2). This COMT \times DARPP-32 interaction effect on d' slope was also present when considering only White participants, $F(2, 64) = 3.18, p = .048$. Although post hoc ANOVA within each COMT group did not reveal statistically significant effects of DARPP-32 genotype, we observed opposing directions for the effects of a putative increase in DRD1 signaling (DARPP-32 T/T) in those with putative low tonic PFC DA (COMT^{Val/Val}) and COMT^{Met} allele carriers. We observed a tendency toward less decrement in target discrimination with increasing load, that is, a less negative d' slope in the COMT^{Val/Val} group, $t(27) = 1.45, p = .16$. In contrast, we observed greater decrement in target discrimination with increasing WM load in those with one, $t(41) = -2.39, p = .022$, or two, $t(23) = -1.48, p = .15$, COMT^{Met} alleles, and when considering COMT^{Met} carriers as a combined group, $t(66) = -2.53, p = .014$. In considering only White participants, the DARPP-32 T/T genotype was associated with significantly less decrement in WM performance with increasing load (less negative d' slope) in COMT^{Val/Val} individuals, $F(1, 16) = 4.13, p = .02$, and nonsignificant increases in the d' slope

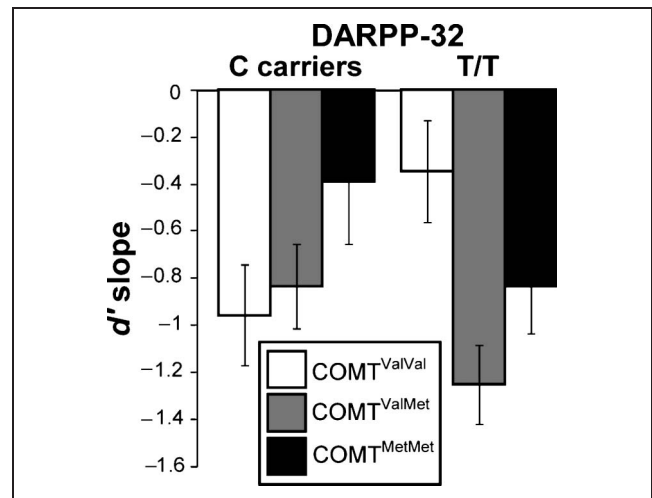


Figure 2. Interacting effects of COMT and DARPP-32 genotype on target discrimination (target/lure d' ; mean \pm SEM) as a function of WM load, as indexed by d' slope, $F(2, 89) = 4.26, p = .017$. The magnitude of d' slope follows an inverted-U pattern, with putatively intermediate levels of frontal DRD1 signaling associated with better performance.

in those with one, $F(1, 29) = 1.76, p = .20$, or two, $F(1, 17) = 0.64, p = .44$, COMT^{Met} alleles. Finally, we directly compared the effect of DARPP-32 genotype on d' slope in COMT heterozygotes and each homozygous group. The effect of DARPP-32 T/T genotype in COMT^{Val/Met} group differed significantly from that in the COMT^{Val/Val} group, $F(1, 66) = 8.17, p = .006$, but did not differ from the COMT^{Met/Met} group, $F(1, 62) = 0.04, p = .85$.

COMT \times DARPP-32 Interaction Effect: Frontal DA or Fronto-striatal DA?

The fact that DARPP-32 boosts DRD1 signaling strongly implicates DARPP-32 in modulating frontal DA-dependent behaviors. Indeed the work of Kolata et al. (2010), Meyer-Lindenberg et al. (2007), and Hotte et al. (2006), all suggest that DARPP-32 modulates frontal function. However, DARPP-32 is most highly expressed in the striatum (Ouimet et al., 1984; Walaas & Greengard, 1984), and the majority of work on DARPP-32 in humans to date has focused on its role in modulating striatal function (Frank & Fossella, 2011; Frank et al., 2007, 2009). Furthermore, striatal DA also plays a role in WM function, where it is thought to primarily subserve successful updating, whereas distractor-resistant maintenance is thought to rely more specifically on frontal DA (see van Schouwenburg et al., 2010). Thus, the interacting effects of DARPP-32 and COMT may not simply reflect effects on frontal DA signaling, but rather interactions between frontal and striatal DA. If the DARPP-32 effect is largely striatal, one would expect an alternative marker of striatal DA to effectively replace it in our statistical models.

To test this possibility, we assayed DAT VNTR genotypes in these participants and repeated our ANOVA for both target/lure discrimination (d') and d' slope, replacing our DARPP-32 genotype factor with DAT genotype: DAT 10/10 repeat individuals ($n = 54$) versus DAT 9 repeat carriers (9R; $n = 40$), with the 9R genotype most frequently associated with enhanced DAT availability and, thus, reduced striatal DA signaling (Costa, Riedel, Muller, Moller, & Ettinger, 2011; Shumay et al., 2011; van de Giessen et al., 2009; van Dyck et al., 2005; Jacobsen et al., 2000). Considering target/lure d' as our dependent measure, in contrast to our significant Load \times COMT \times DARPP-32 interaction, we observed no significant interaction between WM load, COMT, and DAT, $F(2, 86) = 1.85, p = .16$. Likewise, when taking target/lure d' slope as our dependent measure, we observed no significant COMT \times DAT interaction, $F(2, 86) = 1.72, p = .19$. Although not definitive, these findings fail to support the idea that our observed WM Load \times COMT \times DARPP-32 interaction primarily reflects an interaction between frontal and striatal DA tone.

Theoretical Model

Together, our findings are consistent with an inverted-U model of DRD1 signaling, possibly at the level of PFC, on WM task performance under high load (Figure 3), which is expected based on findings in the animal literature (Vijayraghavan et al., 2007). In brief, based on individual putative tonic PFC DA level (COMT Val158Met genotype), greater DRD1 signaling (DARPP-32 T/T genotype) may facilitate, impair, or have no effect on high load WM performance by shifting where an individual falls on the inverted-U curve (see model; Figure 3).

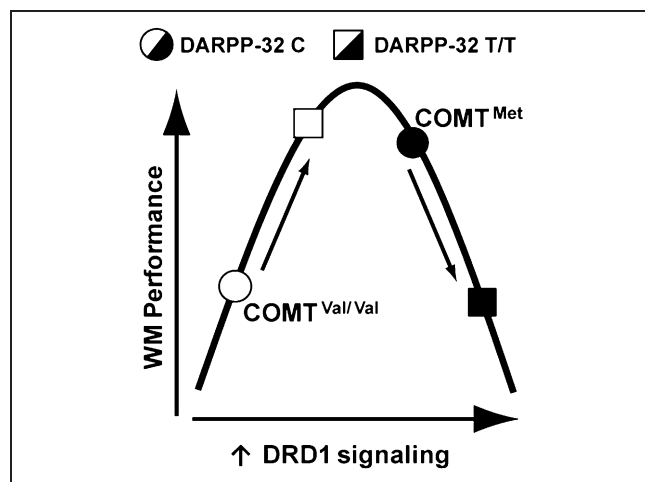


Figure 3. Model depicting an inverted-U relationship of DRD1 signaling, which accounts for interacting effects of COMT and DARPP-32 genetic polymorphisms on WM performance. Intermediate levels of DRD1 signaling, which we propose are acting at the level of PFC, are hypothesized to enable optimal discrimination of target stimuli from interfering stimuli under high WM load.

DISCUSSION

The present findings support an inverted-U model for the role of DA signaling in PFC-dependent WM task performance, in that putatively enhanced DRD1 signaling (DARPP-32 T/T genotype) had differing effects based on putative tonic PFC DA levels (COMT genotype). We found that putatively enhanced DRD1 signaling via the DARPP-32 T allele tends to aid target detection in individuals with presumed low frontal DA tone (COMT^{Val/Val}) but tends to impair target detection in those with putatively higher frontal DA levels (COMT^{Met} carriers). These findings suggest that DRD1 signaling, perhaps largely in PFC, is critical for optimal discrimination of targets from lures when maintenance demands on WM are high. The interpretation that DRD1 signaling is especially important in maintaining target representation in WM amid interfering stimuli is consistent with the demonstrated role of PFC DRD1 signaling in top-down control of visual attention (Noudoost & Moore, 2011).

Prefrontal DRD1 Signaling and WM—Pharmacology Studies

Electrophysiological work in nonhuman primates long ago identified neurons near the principal sulcus (dorsolateral PFC) that sustain firing activity over a delay interval and encode target location information in oculomotor delayed-response tasks (Funahashi, Bruce, & Goldman-Rakic, 1989). Pharmacological studies of these neurons subsequently demonstrated that sustained firing during a delay is critically sensitive to DRD1 manipulations (Sawaguchi, 2001). For example, DRD1 antagonists prevent maintenance of task-relevant information during the delay period in oculomotor delayed-response tasks (Sawaguchi & Goldman-Rakic, 1991, 1994). Furthermore, investigations of both DRD1 agonists and antagonists demonstrated that WM performance is sensitive to DRD1 stimulation according to an inverted-U function, such that either insufficient or excessive DRD1 stimulation degrades performance and that pharmacological effects depend on tonic catecholamine levels (Cai & Arnsten, 1997; Arnsten, Cai, Murphy, & Goldman-Rakic, 1994). Although DRD1-specific agents are not currently approved for use in human participants, available human studies support this inverted-U model for the role of DRD1 in WM processes as DA stimulation enhances WM performance in those with poor baseline WM (Costa, Peppe, Dell’Agnello, Caltagirone, & Carlesimo, 2009; Mehta et al., 2000) and hinders performance in those with high baseline WM capacity (Mattay et al., 2000). Thus, our present findings support the conclusion drawn from pharmacological studies that DRD1 signaling in PFC modulates WM performance according to an inverted-U function.

Prefrontal DA and WM—Genetic Studies

Another approach to understanding the role of neuro-modulators in WM is by evaluating the effect of functional

polymorphisms in genes that regulate signaling in one or more neuromodulatory systems. A rather extensively studied example is the COMT Val158Met polymorphism, which is thought to roughly index tonic PFC DA, a notion supported by recent PET data (Wu et al., 2012). Accounting for tonic PFC DA levels via COMT genotype can help explain the inverted-U effect of pharmacological stimulation of DA signaling on WM performance. For example, the monoamine agonist amphetamine impairs WM in individuals with high tonic PFC DA (COMT^{MetMet}) but enhances WM in individuals with low tonic frontal DA (Mattay et al., 2003). Furthermore, inhibiting COMT improves WM performance in COMT^{ValVal} individuals but worsens performance in COMT^{MetMet} individuals (Farrell, Tunbridge, Braeutigam, & Harrison, 2012). Elevating catecholamine signaling in PFC by inducing stress (Arnsten, 2009) impairs WM performance in COMT^{MetMet} relative to COMT^{ValVal} individuals (Buckert, Kudielka, Reuter, & Fiebach, 2012). Of particular note, Buckert et al. (2012) tested participants in a similar age range to that tested here and used a similar *n*-back task and *d'* index of WM performance. Thus, converging evidence indicates that amplifying DA signaling in PFC interacts with COMT Val158Met genotype according to a unified inverted-U model of PFC DA effects on WM performance.

PFC DRD1 Signaling and Maintaining WM Representations amid Interference

Our findings support the role of putative PFC DRD1 signaling in WM performance following the inverted-U model under specific task demands. In the particular *n*-back task employed here, we found that COMT and DARPP-32 genotype interacted to affect target discrimination only when maintenance demands are high (3-back trials). Importantly, this effect was most pronounced when maintenance of a target representation amid interfering stimuli was considered (i.e., *d'* for correct target detection vs. lure false alarms) as opposed to maintenance of a target among any nontargets, including low interfering nonlures. Thus, the present data suggest that intermediate frontal DRD1 signaling is required for optimal performance specifically in tasks with high maintenance demands under considerable interference. Such a finding is consistent with the proposition that COMT's effect on cognition is most apparent in cognitive tasks which engage the frontal cortical neurobiology tuned by DA (Goldman, Weinberger, Malhotra, & Goldberg, 2009).

Neurocomputational models of PFC function suggest that DRD1 activation enhances persistent high activity (or "up") states thought to underlie the maintenance of information in WM, while depressing spontaneous background activity (Durstewitz & Seamans, 2002). Thus, DRD1 signaling is thought to enhance the "signal" of a WM representation relative to background noise. Importantly, modeling suggests that DRD1-mediated changes in *N*-methyl-D-aspartate and GABA_A currents make persistent activity

in PFC networks less susceptible to interruption by interfering stimuli and enable WM representations robust to interference (Durstewitz, Seamans, & Sejnowski, 2000). Thus, our finding that genetic variations which affect DA tone in PFC presynaptically and DRD1 signaling postsynaptically predict individual differences in the capacity to detect targets under conditions requiring a high WM load within the context of significant interfering stimuli support existing models of PFC DA-dependent WM function.

Study Limitations

Although the present data are consistent with and significantly extend the existing literature regarding DA regulation of WM, we acknowledge the limitations of this study. First, our sample size is relatively small and was thus not adequately powered to detect effects of other polymorphisms that impact DA signaling on WM performance. Although we have assumed a role for the DRD1 modulator DARPP-32 in PFC function, based on the predominance of DRD1 in PFC (Sesack et al., 1995; Goldman-Rakic et al., 1992; Lidow et al., 1991) and the key role for PFC in the maintenance aspect of WM, DARPP-32 has also been implicated in forms of reward learning that depend on striatal DA (Frank et al., 2007, 2009). However, our data lean toward the interpretation that the interacting effects of COMT and DARPP-32 on high load WM performance reflects PFC DRD1 signaling, as when we replaced DARPP-32 genotype in our models with a DAT VNTR genotype associated with variation in striatal DA tone, we found no significant interaction. Even considering these negative DAT results, however, we cannot rule out the possibility that our results here reflect an interaction between frontal and striatal DA, perhaps via striatal gating of goal-related information into WM at the level of PFC (van Schouwenburg et al., 2010; Gruber, Dayan, Gutkin, & Solla, 2006; Frank, Loughry, & O'Reilly, 2001). Future PET studies quantifying striatal and extrastriatal DA signaling as a function of genotype coupled with measures of WM performance will allow for a more definitive examination of this issue. Finally, it is important to note that COMT and DARPP-32 modulate both DRD1- and DRD2-mediated signaling, albeit with different downstream effects in the case of DARPP-32 (Greengard et al., 1999). However, based on evidence demonstrating the critical role of DRD1 signaling localized to PFC in WM maintenance (Vijayraghavan et al., 2007; Goldman-Rakic et al., 2000; Sawaguchi & Goldman-Rakic, 1991), it seems parsimonious to interpret our COMT × DARPP-32 interaction effect as reflecting DRD1-mediated WM maintenance processes in PFC. Moreover, we observed this effect on 3-back but not 2-back trials, which differ only in the number of items that must be maintained in memory to correctly identify targets. Nonetheless, further dissection of the roles of DRD1 and DRD2 signaling in these WM processes is warranted.

Dopaminergic Functioning Varies with Age

With regard to the role of COMT in cognition, not all of the published literature agrees. The present data argue that DARPP-32 variation could underlie some differences within COMT genotype. However, developmental changes in frontal DA neurotransmission (Wahlstrom, Collins, White, & Luciana, 2010; Tunbridge et al., 2007) likely interacts with genetic effects on cognition. Such effects may contribute to conflicting results from studies investigating the role of COMT in executive function. For instance, the participants in Mattay et al. (2003) were in their mid thirties, whereas participants in a recent study that failed to replicate Mattay et al.'s (2003) effects had a mean age of 23 years (Wardle, Hart, Palmer, & de Wit, 2013). Whereas the current study focused on participants aged 22–40 years to limit age-related variability in frontal DA signaling, whether similar genotype effects on WM are present among the frequent participants of cognitive studies, emerging adults, remains an open question.

Conclusion

Our results are consistent with existing computational models for the role of PFC DRD1 signaling in WM processes (Durstewitz & Seamans, 2002). Specifically, we have shown that putative DRD1 signaling (indexed by COMT and DARPP-32 genotype) predicts the stability of target representation in the face of interfering stimuli according to an inverted-U function (Cools & D'Esposito, 2011). Furthermore, our findings support the conclusion that inverted-U-shaped DA actions on human WM and cognitive control may be particularly strong under conditions of high cognitive load.

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REFERENCES

Anchordoquy, H. C., McGeary, C., Liu, L., Krauter, K. S., & Smolen, A. (2003). Genotyping of three candidate genes after whole-genome preamplification of DNA collected from buccal cells. *Behavior Genetics*, *33*, 73–78.

Arnsten, A. F. (2009). Stress signalling pathways that impair prefrontal cortex structure and function. *Nature Reviews Neuroscience*, *10*, 410–422.

Arnsten, A. F., Cai, J. X., Murphy, B. L., & Goldman-Rakic, P. S. (1994). Dopamine D1 receptor mechanisms in the cognitive performance of young adult and aged monkeys. *Psychopharmacology (Berlin)*, *116*, 143–151.

Barnett, J. H., Scoriels, L., & Munafo, M. R. (2008). Meta-analysis of the cognitive effects of the catechol-O-methyltransferase gene Val158/108Met polymorphism. *Biological Psychiatry*, *64*, 137–144.

Bilder, R. M., Volavka, J., Lachman, H. M., & Grace, A. A. (2004). The catechol-O-methyltransferase polymorphism: Relations to the tonic-phasic dopamine hypothesis and neuropsychiatric phenotypes. *Neuropsychopharmacology*, *29*, 1943–1961.

Boettiger, C. A., Mitchell, J. M., Tavares, V. C., Robertson, M., Joslyn, G., D'Esposito, M., et al. (2007). Immediate reward bias in humans: Fronto-parietal networks and a role for the catechol-O-methyltransferase 158(Val/Val) genotype. *Journal of Neuroscience*, *27*, 14383–14391.

Bruder, G. E., Keilp, J. G., Xu, H., Shikhman, M., Schori, E., Gorman, J. M., et al. (2005). Catechol-O-methyltransferase (COMT) genotypes and working memory: Associations with differing cognitive operations. *Biological Psychiatry*, *58*, 901–907.

Buckert, M., Kudielka, B. M., Reuter, M., & Fiebich, C. J. (2012). The COMT Val158Met polymorphism modulates working memory performance under acute stress. *Psychoneuroendocrinology*, *37*, 1810–1821.

Cai, J. X., & Arnsten, A. F. (1997). Dose-dependent effects of the dopamine D1 receptor agonists A77636 or SKF81297 on spatial working memory in aged monkeys. *Journal of Pharmacology and Experimental Therapeutics*, *283*, 183–189.

Chen, J., Lipska, B. K., Halim, N., Ma, Q. D., Matsumoto, M., Melhem, S., et al. (2004). Functional analysis of genetic variation in catechol-O-methyltransferase (COMT): Effects on mRNA, protein, and enzyme activity in postmortem human brain. *The American Journal of Human Genetics*, *75*, 807–821.

Cools, R., & D'Esposito, M. (2011). Inverted-U-shaped dopamine actions on human working memory and cognitive control. *Biological Psychiatry*, *69*, e113–e125.

Cools, R., Gibbs, S. E., Miyakawa, A., Jagust, W., & D'Esposito, M. (2008). Working memory capacity predicts dopamine synthesis capacity in the human striatum. *Journal of Neuroscience*, *28*, 1208–1212.

Costa, A., Peppe, A., Dell'Agnello, G., Caltagirone, C., & Carlesimo, G. A. (2009). Dopamine and cognitive functioning in de novo subjects with Parkinson's disease: Effects of pramipexole and pergolide on working memory. *Neuropsychologia*, *47*, 1374–1381.

Costa, A., Riedel, M., Muller, U., Moller, H. J., & Ettinger, U. (2011). Relationship between SLC6A3 genotype and striatal dopamine transporter availability: A meta-analysis of human single photon emission computed tomography studies. *Synapse*, *65*, 998–1005.

Dennis, N. A., Need, A. C., LaBar, K. S., Waters-Metenier, S., Cirulli, E. T., Kragel, J., et al. (2010). COMT val108/158 met genotype affects neural but not cognitive processing in healthy individuals. *Cerebral Cortex*, *20*, 672–683.

Dosenbach, N. U., Nardos, B., Cohen, A. L., Fair, D. A., Power, J. D., Church, J. A., et al. (2010). Prediction of individual brain maturity using fMRI. *Science*, *329*, 1358–1361.

Durstewitz, D., & Seamans, J. K. (2002). The computational role of dopamine D1 receptors in working memory. *Neural Networks*, *15*, 561–572.

Durstewitz, D., & Seamans, J. K. (2008). The dual-state theory of prefrontal cortex dopamine function with relevance to catechol-O-methyltransferase genotypes and schizophrenia. *Biological Psychiatry*, *64*, 739–749.

Durstewitz, D., Seamans, J. K., & Sejnowski, T. J. (2000). Dopamine-mediated stabilization of delay-period activity

- in a network model of prefrontal cortex. *Journal of Neurophysiology*, *83*, 1733–1750.
- Durstewitz, D., Vitoz, N. M., Floresco, S. B., & Seamans, J. K. (2010). Abrupt transitions between prefrontal neural ensemble states accompany behavioral transitions during rule learning. *Neuron*, *66*, 438–448.
- Egan, M. F., Goldberg, T. E., Kolachana, B. S., Callicott, J. H., Mazzanti, C. M., Straub, R. E., et al. (2001). Effect of COMT Val108/158 Met genotype on frontal lobe function and risk for schizophrenia. *Proceedings of the National Academy of Sciences, U.S.A.*, *98*, 6917–6922.
- Farrell, S. M., Tunbridge, E. M., Braeutigam, S., & Harrison, P. J. (2012). COMT Val(158)Met genotype determines the direction of cognitive effects produced by catechol-O-methyltransferase inhibition. *Biological Psychiatry*, *71*, 538–544.
- Floresco, S. B., & Phillips, A. G. (2001). Delay-dependent modulation of memory retrieval by infusion of a dopamine D1 agonist into the rat medial prefrontal cortex. *Behavioral Neuroscience*, *115*, 934–939.
- Frank, M. J., Doll, B. B., Oas-Terpstra, J., & Moreno, F. (2009). Prefrontal and striatal dopaminergic genes predict individual differences in exploration and exploitation. *Nature Neuroscience*, *12*, 1062–1068.
- Frank, M. J., & Fossella, J. A. (2011). Neurogenetics and pharmacology of learning, motivation, and cognition. *Neuropsychopharmacology*, *36*, 133–152.
- Frank, M. J., Loughry, B., & O'Reilly, R. C. (2001). Interactions between frontal cortex and basal ganglia in working memory: A computational model. *Cognitive, Affective & Behavioral Neuroscience*, *1*, 137–160.
- Frank, M. J., Moustafa, A. A., Haughey, H. M., Curran, T., & Hutchison, K. E. (2007). Genetic triple dissociation reveals multiple roles for dopamine in reinforcement learning. *Proceedings of the National Academy of Sciences, U.S.A.*, *104*, 16311–16316.
- Frazer, K. A., Ballinger, D. G., Cox, D. R., Hinds, D. A., Stuve, L. L., Gibbs, R. A., et al. (2007). A second generation human haplotype map of over 3.1 million SNPs. *Nature*, *449*, 851–861.
- Funahashi, S., Bruce, C. J., & Goldman-Rakic, P. S. (1989). Mnemonic coding of visual space in the monkey's dorsolateral prefrontal cortex. *Journal of Neurophysiology*, *61*, 331–349.
- Gaspar, P., Bloch, B., & Le Moine, C. (1995). D1 and D2 receptor gene expression in the rat frontal cortex: Cellular localization in different classes of efferent neurons. *European Journal of Neuroscience*, *7*, 1050–1063.
- Giros, B., Jaber, M., Jones, S. R., Wightman, R. M., & Caron, M. G. (1996). Hyperlocomotion and indifference to cocaine and amphetamine in mice lacking the dopamine transporter. *Nature*, *379*, 606–612.
- Gogos, J. A., Morgan, M., Luine, V., Santha, M., Ogawa, S., Pfaff, D., et al. (1998). Catechol-O-methyltransferase-deficient mice exhibit sexually dimorphic changes in catecholamine levels and behavior. *Proceedings of the National Academy of Sciences, U.S.A.*, *95*, 9991–9996.
- Goldberg, T. E., Egan, M. F., Gscheidle, T., Coppola, R., Weickert, T., Kolachana, B. S., et al. (2003). Executive subprocesses in working memory: Relationship to catechol-O-methyltransferase Val158Met genotype and schizophrenia. *Archives of General Psychiatry*, *60*, 889–896.
- Goldman, D., Weinberger, D. R., Malhotra, A. K., & Goldberg, T. E. (2009). The role of COMT Val158Met in cognition. *Biological Psychiatry*, *65*, e1–e2; author reply e3–4.
- Goldman-Rakic, P. S. (1996). The prefrontal landscape: Implications of functional architecture for understanding human mentation and the central executive. *Philosophical Transactions of the Royal Society of London, Series B, Biological Sciences*, *351*, 1445–1453.
- Goldman-Rakic, P. S., Lidow, M. S., Smiley, J. F., & Williams, M. S. (1992). The anatomy of dopamine in monkey and human prefrontal cortex. *Journal of Neural Transmission, Supplementum*, *36*, 163–177.
- Goldman-Rakic, P. S., Muly, E. C., III, & Williams, G. V. (2000). D(1) receptors in prefrontal cells and circuits. *Brain Research. Brain Research Reviews*, *31*, 295–301.
- Gray, J. R., Chabris, C. F., & Braver, T. S. (2003). Neural mechanisms of general fluid intelligence. *Nature Neuroscience*, *6*, 316–322.
- Green, D., & Swets, J. (1966). *Signal detection theory and psychophysics*. New York: Wiley.
- Greengard, P., Allen, P. B., & Nairn, A. C. (1999). Beyond the dopamine receptor: The DARPP-32/protein phosphatase-1 cascade. *Neuron*, *23*, 435–447.
- Gruber, A. J., Dayan, P., Gutkin, B. S., & Solla, S. A. (2006). Dopamine modulation in the basal ganglia locks the gate to working memory. *Journal of Computational Neuroscience*, *20*, 153–166.
- Heinz, A., Goldman, D., Jones, D. W., Palmour, R., Hommer, D., Gorey, J. G., et al. (2000). Genotype influences in vivo dopamine transporter availability in human striatum. *Neuropsychopharmacology*, *22*, 133–139.
- Heyser, C. J., Fienberg, A. A., Greengard, P., & Gold, L. H. (2000). DARPP-32 knockout mice exhibit impaired reversal learning in a discriminated operant task. *Brain Research*, *867*, 122–130.
- Hollingshead, A. (1975). *Hollingshead's Four Factor Index of Social Status*. New Haven, CT: Yale University Press.
- Hotte, M., Thuault, S., Lachaise, F., Dineley, K. T., Hemmings, H. C., Nairn, A. C., et al. (2006). D1 receptor modulation of memory retrieval performance is associated with changes in pCREB and pDARPP-32 in rat prefrontal cortex. *Behavioural Brain Research*, *171*, 127–133.
- Jacobs, E., & D'Esposito, M. (2011). Estrogen shapes dopamine-dependent cognitive processes: Implications for women's health. *Journal of Neuroscience*, *31*, 5286–5293.
- Jacobsen, L. K., Staley, J. K., Zoghbi, S. S., Seibyl, J. P., Kosten, T. R., Innis, R. B., et al. (2000). Prediction of dopamine transporter binding availability by genotype: A preliminary report. *American Journal of Psychiatry*, *157*, 1700–1703.
- Jones, S. R., Gainetdinov, R. R., Wightman, R. M., & Caron, M. G. (1998). Mechanisms of amphetamine action revealed in mice lacking the dopamine transporter. *Journal of Neuroscience*, *18*, 1979–1986.
- Kaenmaki, M., Tammimaki, A., Myohanen, T., Pakarinen, K., Amberg, C., Karayiorgou, M., et al. (2010). Quantitative role of COMT in dopamine clearance in the prefrontal cortex of freely moving mice. *Journal of Neurochemistry*, *114*, 1745–1755.
- Kang, A. M., Palmatier, M. A., & Kidd, K. K. (1999). Global variation of a 40-bp VNTR in the 3'-untranslated region of the dopamine transporter gene (SLC6A3). *Biological Psychiatry*, *46*, 151–160.
- Karoum, F., Chrapusta, S. J., & Egan, M. F. (1994). 3-Methoxytyramine is the major metabolite of released dopamine in the rat frontal cortex: Reassessment of the effects of antipsychotics on the dynamics of dopamine release and metabolism in the frontal cortex, nucleus accumbens, and striatum by a simple two pool model. *Journal of Neurochemistry*, *63*, 972–979.
- Kolata, S., Light, K., Wass, C. D., Colas-Zelin, D., Roy, D., & Matzel, L. D. (2010). A dopaminergic gene cluster in the prefrontal cortex predicts performance indicative of

- general intelligence in genetically heterogeneous mice. *PLoS One*, *5*, e14036.
- Kroener, S., Chandler, L. J., Phillips, P. E., & Seamans, J. K. (2009). Dopamine modulates persistent synaptic activity and enhances the signal-to-noise ratio in the prefrontal cortex. *PLoS One*, *4*, e6507.
- Landau, S. M., Lal, R., O'Neil, J. P., Baker, S., & Jagust, W. J. (2009). Striatal dopamine and working memory. *Cerebral Cortex*, *19*, 445–454.
- Lewis, D. A., Melchitzky, D. S., Sesack, S. R., Whitehead, R. E., Auh, S., & Sampson, A. (2001). Dopamine transporter immunoreactivity in monkey cerebral cortex: Regional, laminar, and ultrastructural localization. *Journal of Comparative Neurology*, *432*, 119–136.
- Lidow, M. S., Goldman-Rakic, P. S., Gallager, D. W., & Rakic, P. (1991). Distribution of dopaminergic receptors in the primate cerebral cortex: Quantitative autoradiographic analysis using [³H]raclopride, [³H]spiperone and [³H]SCH23390. *Neuroscience*, *40*, 657–671.
- Lidow, M. S., Koh, P. O., & Arnsten, A. F. (2003). D1 dopamine receptors in the mouse prefrontal cortex: Immunocytochemical and cognitive neuropharmacological analyses. *Synapse*, *47*, 101–108.
- Macmillan, N., & Creelman, C. (2005). *Signal detection theory: A user's guide* (2nd ed.). Mahwah, NJ: Lawrence Erlbaum Associates.
- Malhotra, A. K., Kestler, L. J., Mazzanti, C., Bates, J. A., Goldberg, T., & Goldman, D. (2002). A functional polymorphism in the COMT gene and performance on a test of prefrontal cognition. *American Journal of Psychiatry*, *159*, 652–654.
- Mattay, V. S., Callicott, J. H., Bertolino, A., Heaton, I., Frank, J. A., Coppola, R., et al. (2000). Effects of dextroamphetamine on cognitive performance and cortical activation. *NeuroImage*, *12*, 268–275.
- Mattay, V. S., Goldberg, T. E., Fera, F., Hariri, A. R., Tessitore, A., Egan, M. F., et al. (2003). Catechol O-methyltransferase val158-met genotype and individual variation in the brain response to amphetamine. *Proceedings of the National Academy of Sciences, U.S.A.*, *100*, 6186–6191.
- McLeod, H. L., Fang, L., Luo, X., Scott, E. P., & Evans, W. E. (1994). Ethnic differences in erythrocyte catechol-O-methyltransferase activity in Black and White Americans. *Journal of Pharmacology and Experimental Therapeutics*, *270*, 26–29.
- McLeod, H. L., Syvanen, A. C., Githang'a, J., Indalo, A., Ismail, D., Dewar, K., et al. (1998). Ethnic differences in catechol-O-methyltransferase pharmacogenetics: Frequency of the codon 108/158 low activity allele is lower in Kenyan than Caucasian or South-west Asian individuals. *Pharmacogenetics*, *8*, 195–199.
- Mehta, M. A., Owen, A. M., Sahakian, B. J., Mavaddat, N., Pickard, J. D., & Robbins, T. W. (2000). Methylphenidate enhances working memory by modulating discrete frontal and parietal lobe regions in the human brain. *Journal of Neuroscience*, *20*, RC65.
- Meyer-Lindenberg, A., Straub, R. E., Lipska, B. K., Verchinski, B. A., Goldberg, T., Callicott, J. H., et al. (2007). Genetic evidence implicating DARPP-32 in human frontostriatal structure, function, and cognition. *The Journal of Clinical Investigation*, *117*, 672–682.
- Mitchell, R. J., Howlett, S., Earl, L., White, N. G., McComb, J., Schanfield, M. S., et al. (2000). Distribution of the 3' VNTR polymorphism in the human dopamine transporter gene in world populations. *Human Biology*, *72*, 295–304.
- Nishi, A., Snyder, G. L., & Greengard, P. (1997). Bidirectional regulation of DARPP-32 phosphorylation by dopamine. *Journal of Neuroscience*, *17*, 8147–8155.
- Noudoost, B., & Moore, T. (2011). Control of visual cortical signals by prefrontal dopamine. *Nature*, *474*, 372–375.
- Ouimet, C. C., Miller, P. E., Hemmings, H. C., Jr., Walaas, S. I., & Greengard, P. (1984). DARPP-32, a dopamine- and adenosine 3':5'-monophosphate-regulated phosphoprotein enriched in dopamine-innervated brain regions. III. Immunocytochemical localization. *Journal of Neuroscience*, *4*, 111–124.
- Rieckmann, A., Karlsson, S., Fischer, H., & Backman, L. (2011). Caudate dopamine D1 receptor density is associated with individual differences in frontoparietal connectivity during working memory. *Journal of Neuroscience*, *31*, 14284–14290.
- Sawaguchi, T. (2001). The effects of dopamine and its antagonists on directional delay-period activity of prefrontal neurons in monkeys during an oculomotor delayed-response task. *Neuroscience Research*, *41*, 115–128.
- Sawaguchi, T., & Goldman-Rakic, P. S. (1991). D1 dopamine receptors in prefrontal cortex: Involvement in working memory. *Science*, *251*, 947–950.
- Sawaguchi, T., & Goldman-Rakic, P. S. (1994). The role of D1-dopamine receptor in working memory: Local injections of dopamine antagonists into the prefrontal cortex of rhesus monkeys performing an oculomotor delayed-response task. *Journal of Neurophysiology*, *71*, 515–528.
- Seamans, J. K., Floresco, S. B., & Phillips, A. G. (1998). D1 receptor modulation of hippocampal-prefrontal cortical circuits integrating spatial memory with executive functions in the rat. *Journal of Neuroscience*, *18*, 1613–1621.
- Seamans, J. K., & Yang, C. R. (2004). The principal features and mechanisms of dopamine modulation in the prefrontal cortex. *Progress in Neurobiology*, *74*, 1–58.
- Sesack, S. R., Hawrylak, V. A., Matus, C., Guido, M. A., & Levey, A. I. (1998). Dopamine axon varicosities in the prelimbic division of the rat prefrontal cortex exhibit sparse immunoreactivity for the dopamine transporter. *Journal of Neuroscience*, *18*, 2697–2708.
- Sesack, S. R., Snyder, C. L., & Lewis, D. A. (1995). Axon terminals immunolabeled for dopamine or tyrosine hydroxylase synapse on GABA-immunoreactive dendrites in rat and monkey cortex. *Journal of Comparative Neurology*, *363*, 264–280.
- Shumay, E., Chen, J., Fowler, J. S., & Volkow, N. D. (2011). Genotype and ancestry modulate brain's DAT availability in healthy humans. *PLoS One*, *6*, e22754.
- Smith, C. T., & Boettiger, C. A. (2012). Age modulates the effect of COMT genotype on delay discounting behavior. *Psychopharmacology (Berlin)*, *222*, 609–617.
- Stelzel, C., Basten, U., Montag, C., Reuter, M., & Fiebach, C. J. (2010). Frontostriatal involvement in task switching depends on genetic differences in d2 receptor density. *Journal of Neuroscience*, *30*, 14205–14212.
- Svenningsson, P., Nishi, A., Fisone, G., Girault, J. A., Nairn, A. C., & Greengard, P. (2004). DARPP-32: An integrator of neurotransmission. *Annual Review of Pharmacology and Toxicology*, *44*, 269–296.
- Tunbridge, E. M., Bannerman, D. M., Sharp, T., & Harrison, P. J. (2004). Catechol-o-methyltransferase inhibition improves set-shifting performance and elevates stimulated dopamine release in the rat prefrontal cortex. *Journal of Neuroscience*, *24*, 5331–5335.
- Tunbridge, E. M., Weickert, C. S., Kleinman, J. E., Herman, M. M., Chen, J., Kolachana, B. S., et al. (2007). Catechol-o-methyltransferase enzyme activity and protein expression in human prefrontal cortex across the postnatal lifespan. *Cerebral Cortex*, *17*, 1206–1212.
- van de Giessen, E., de Win, M. M., Tanck, M. W., van den Brink, W., Baas, F., & Booij, J. (2009). Striatal dopamine transporter

- availability associated with polymorphisms in the dopamine transporter gene SLC6A3. *Journal of Nuclear Medicine*, 50, 45–52.
- van Dyck, C. H., Malison, R. T., Jacobsen, L. K., Seibyl, J. P., Staley, J. K., Laruelle, M., et al. (2005). Increased dopamine transporter availability associated with the 9-repeat allele of the SLC6A3 gene. *Journal of Nuclear Medicine*, 46, 745–751.
- van Schouwenburg, M., Aarts, E., & Cools, R. (2010). Dopaminergic modulation of cognitive control: Distinct roles for the prefrontal cortex and the basal ganglia. *Current Pharmaceutical Design*, 16, 2026–2032.
- Vandenbergh, D. J., Persico, A. M., Hawkins, A. L., Griffin, C. A., Li, X., Jabs, E. W., et al. (1992). Human dopamine transporter gene (DAT1) maps to chromosome 5p15.3 and displays a VNTR. *Genomics*, 14, 1104–1106.
- VanNess, S. H., Owens, M. J., & Kilts, C. D. (2005). The variable number of tandem repeats element in DAT1 regulates in vitro dopamine transporter density. *BMC Genetics*, 6, 55.
- Vijayraghavan, S., Wang, M., Birnbaum, S. G., Williams, G. V., & Arnsten, A. F. (2007). Inverted-U dopamine D1 receptor actions on prefrontal neurons engaged in working memory. *Nature Neuroscience*, 10, 376–384.
- Wahlstrom, D., Collins, P., White, T., & Luciana, M. (2010). Developmental changes in dopamine neurotransmission in adolescence: Behavioral implications and issues in assessment. *Brain and Cognition*, 72, 146–159.
- Walaas, S. I., & Greengard, P. (1984). DARPP-32, a dopamine- and adenosine 3':5'-monophosphate-regulated phosphoprotein enriched in dopamine-innervated brain regions. I. Regional and cellular distribution in the rat brain. *Journal of Neuroscience*, 4, 84–98.
- Wardle, M. C., Hart, A. B., Palmer, A. A., & de Wit, H. (2013). Does COMT genotype influence the effects of d-amphetamine on executive functioning? *Genes, Brain, and Behavior*, 12, 13–20.
- Williams, G. V., & Castner, S. A. (2006). Under the curve: Critical issues for elucidating D1 receptor function in working memory. *Neuroscience*, 139, 263–276.
- Wu, K., O'Keefe, D., Politis, M., O'Keefe, G. C., Robbins, T. W., Bose, S. K., et al. (2012). The catechol-O-methyltransferase Val(158)Met polymorphism modulates fronto-cortical dopamine turnover in early Parkinson's disease: A PET study. *Brain*, 135, 2449–2457.
- Yavich, L., Forsberg, M. M., Karayiorgou, M., Gogos, J. A., & Mannisto, P. T. (2007). Site-specific role of catechol-O-methyltransferase in dopamine overflow within prefrontal cortex and dorsal striatum. *Journal of Neuroscience*, 27, 10196–10209.